ENGINEERS / SCIENTISTS / PROGRAM MANAGERS



March 7, 2021

Mr. Alex Baylor
Environmental Specialist
Environmental Safety Office
Prince George's County Public Schools
Division of Supporting Services / Building Services
13306 Old Marlboro Pike
Upper Marlboro, MD 20772

via email: alex.baylor@pgcps.org

RE: Indoor Air Quality (IAQ) and Mold Assessment Services

Prince George's County Public Schools (PGCPS) – Cooper Lane Elementary School

3817 Cooper Lane, Landover Hills, Maryland 20784

Contract No.: IFB 022-19: Indoor Air Quality Services at Various Locations

Tidewater Project No.: 5419-029

Dear Mr. Baylor:

Tidewater, Inc. (Tidewater) is pleased to present this final report regarding the results of the Indoor Air Quality (IAQ) and Mold Assessment Services conducted by Tidewater at Cooper Lane Elementary School located at 3817 Cooper Lane in Landover Hills, Maryland. Tidewater's Project Manager and Certified Industrial Hygienist, Mr. Skanda Abeyesekere MS, CIH, CSP, CHMM, conducted these services on November 19, 2020. Re-sampling of areas with elevated mold concentrations were conducted on March 2, 2021.

The scope of work for the IAQ assessment and mold survey included:

- The following typical occupied areas of the school chosen at the industrial hygienist's discretion for inspection and sampling: Main Office, Library, Classroom 10, Classroom 5, Classroom 2, Classroom 14, Classroom 17, Classroom 25, Health Unit and Multipurpose Room. These areas were inspected for evidence of potential indoor air quality problems (including suspect microbial growth, water damage, chemical use/ storage, drain traps, sources of allergens/ contaminants, etc.) that may contribute to indoor air quality problems;
- Direct read measurements for temperature (T), relative humidity (RH), carbon dioxide (CO₂), and carbon monoxide (CO) in the above locations for comparison with standards established by the American Society of Heating, Refrigerating and Air-Conditioning Engineers (ASHRAE) Standard 62.1–2019, Ventilation for Acceptable Indoor Air Quality, and The United States Environmental Protection Agency (US EPA) National Ambient Air Quality Standards (NAAQS);
- Direct read measurements for Particulate Matter less than 10 microns (PM10) in the above locations for comparison with standards established by the US EPA NAAQS Final Action (December 7, 2020); and
- Air sampling for microbial spores in the above locations for total airborne fungal spore analysis.



Visual Observation

Due to the on-going COID-19 pandemic, the school building was occupied by limited number of staff and no students were present at the time of the survey. As a result, the majority of the classrooms and other common areas inspected were vacant. Tidewater's assessment included a visual inspection of the following areas of Cooper Lane Elementary School chosen at the Industrial hygienist's discretion. The results of Tidewater's visual inspection are as follows:

Main Office

The Main Office appeared to be clean and well maintained. Housekeeping appeared to be satisfactory. No signs of ongoing water-intrusion problems were observed and no odors were detected. One (1) wall-mounted fan coil unit was in operation and was emitting warm air at the time of the inspection. One (1) window-mounted air conditioning unit was also observed.

Library

Two (2) window-mounted air conditioning units were observed in the Library. None of these units were not in operation at the time of the inspection. Multiple wall-mounted fan coil units were in operation and were emitting warm air at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the library and no odors were detected. The library appeared to be well maintained and organized.

Classroom 10

A wall-mounted fan coil unit was in operation and was emitting warm air at the time of the inspection. No signs of mold growth or past or ongoing water-intrusion problems were observed in Classroom 10. Furthermore, no notable odors were detected. The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup. The classroom appeared to be clean.

Classroom 5

One (1) window-mounted air conditioning unit and multiple wall-mounted fan coil units were observed in the classroom. The wall-mounted fan coil units were in operation and were emitting warm air at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the classroom and no odors were detected. The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup.

Classroom 14

Two (2) window-mounted air conditioning units were installed in the classroom. The front panel of one of these units was dismantled and appeared to be broken. Multiple wall-mounted fan coil units were in operation and were emitting warm air at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the classroom and no odors were detected. The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup.

Classroom 17

One (1) window-mounted air conditioning unit and multiple wall-mounted fan coil unit were observed in the classroom. The wall-mounted fan coil unit were in operation and were emitting warm air at the time of the inspection. The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup. The wall-mounted air supply grills appeared to be clean.



No signs of ongoing water-intrusion problems were observed. Furthermore, no odors were detected.

Classroom 25

Two (2) floor-mounted air conditioning units were operating and were emitting warm air at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the classroom and no odors were detected. The classroom appeared to be clean and organized.

Classroom 2

A window-mounted air conditioning unit and multiple wall-mounted fan coil units were observed in the classroom. The wall-mounted fan coil units were in operation and were emitting warm air at the time of the inspection. The supply grills of the window-mounted air conditioning unit appeared to be dusty. The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup. The wall-mounted air supply grills also appeared to be dusty. No signs of ongoing water-intrusion problems were observed in the classroom and no odors were detected.

Health Unit

One (1) window-mounted air conditioning unit was observed in the health unit and was labelled <u>"out of order".</u> The fan coil unit was not in operation at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the health unit and no odors were detected.

Multipurpose Room

Six (6) ceiling-mounted exhaust fans were in operation at the time of the inspection. The multipurpose room was also equipped with four (4) window-mounted air conditioning units which were not in operation at the time of the inspection. No signs of ongoing water-intrusion problems were observed in the multipurpose room and no odors were detected. The ceiling-mounted air supply grills appeared to have dust accumulations and rust buildup.

Comfort Parameter Air Testing

During the assessment, Tidewater obtained temperature (T), relative humidity (RH), carbon dioxide (CO₂), and carbon monoxide (CO) measurements within select locations of the school using a TSI VelociCalc Indoor Air Quality instrument (Model Number 9565-X, Serial Number 9565X 1945 002, Calibration Date: November 8, 2019.) Measurements were taken after allowing the instrument to become acclimated to the ambient temperature and relative humidity for approximately five (5) minutes. Measurements were taken over a 5-minute time period at each designated location and the average concentration was recorded. Samples were obtained for comparison with standards established by the American Society for Heating Refrigeration and Air Conditioning (ASHRAE) Standard 62.1 – 2019, Ventilation for Acceptable Indoor Air Quality. Tidewater also obtained a background sample outdoors in front of the main entrance of the school building for comparison to the interior readings. The results of the IAQ comfort parameter monitoring are provided in Table 1, in **Attachment A.**

According to the American Society for Heating, Refrigerating and Air Conditioning Engineers (ASHRAE) Standard 62.1 – 2019, *Ventilation for Acceptable Indoor Air Quality*, the temperature range in summer months should be maintained between 73.0°F and 79.0°F for maximum occupant comfort. The ASHRAE standard for temperature for winter months is between 68.0°F and 74.5°F. The indoor temperature levels within the assessed areas on November 19, 2020



ranged between 57.8°F and 73.1°F. The background temperature outside the building was 52.1°F. The temperature levels recorded within most areas monitored were within temperature levels typically observed during the fall-winter transitional period. The temperature levels recorded within classroom 25, health unit and the multipurpose room were below the ASHRAE lower temperature standard of 68.0°F recommended for winter months. These areas were vacant at the time of the inspection. Indoor temperature levels tend to fluctuate throughout the work day based on the number of occupants present within the individual work spaces. The temperature level in these areas are likely to be within ASHRAE standards when they are re-occupied.

Per the same ASHRAE standard, a maximum recommended relative humidity level of 65.0% or below is recommended to reduce the likelihood of condensation on cold surfaces. Relative humidity levels within the assessed areas on November 19, 2020 ranged between 20.8% and 42.4%. The background relative humidity level outside the building was 28.9%. The relative humidity levels in all areas assessed were below the ASHRAE recommended maximum relative humidity standard of 65.0%.

ASHRAE Standard 62.1 - 2019 recommends that indoor CO_2 levels not exceed 700 ppm above the outdoor background CO_2 level. The CO_2 levels in the assessed areas on November 19, 2020 ranged between 447 ppm to 578 ppm. The background CO_2 level outside the building was 440 ppm. The CO_2 levels within all interior locations assessed did not exceed 700 ppm above the outdoor background CO_2 level of 440 ppm.

The CO levels in all areas assessed on November 19, 2020 were below the maximum standard of 9.0 ppm recommended by the Indoor Air Quality Association (IAQA) for CO in occupied indoor environments.

Particulate Matter Less Than 10 microns (PM10)

During the assessment, Tidewater obtained particulate matter less than 10 microns (PM10) dust particulate measurements within select locations of the school using a TSI® DUST TRAK IITM Aerosol Monitor (Model 8534, Serial Number 8534170101.) Measurements were taken after allowing the device to become acclimated to the ambient temperature and relative humidity for five (5) minutes. Measurements were taken over a 5-minute time period at each sampling location and the average concentration was recorded for comparison with standards established by the US EPA NAAQS Final Action (December 7, 2020.)

Tidewater also obtained a background sample outdoors in front of the main entrance of the school building for comparison to the interior readings.

The results of the particulate matter sampling are provided in Table 2, in Attachment A.

Based on the EPA NAAQS for Particulate Matter, Final Action (December 7, 2020), the 24-hour primary and secondary exposure standard for particulate matter less than 10 microns (PM10) is 150.0 micrograms per cubic meter of air (μ g/m³) or 0.150 milligrams per cubic meter of air (μ g/m³.) The results of the PM10 analysis indicate that the average PM10 dust concentrations in all assessed areas ranged between 0.071 mg/m³ and 0.083 mg/m³. The average PM10 dust concentration in the background sample obtained in front of the main entrance was 0.074 mg/m³. The PM10 concentrations all areas assessed were below the EPA 24-hour primary and secondary NAAQS of 0.150 mg/m³.



Spore Trap Bioaerosol Sampling

Tidewater collected spore trap air samples from select locations within the school chosen by the Industrial hygienist's discretion to characterize air quality for total airborne total fungal spores. The samples were collected from the same locations where the comfort parameters were recorded. Tidewater obtained the spore trap samples using Allergenco-D cassettes affixed to a Buck BioAire™ Bioaerosol Sampling Pump (Pump Model Number B520 and Serial Number B153043) calibrated to a flow rate of 15.0 Liters per minute. Each sample was run for a period of five (5) minutes at each sample location to collect a total sample volume of 75.0 liters of air. Tidewater also obtained a background sample outdoors in front of the main entrance of the school building for comparison to the interior readings.

Once collected, the samples were transported to EMSL Analytical Laboratory (EMSL) located in Beltsville, Maryland for analysis via a standard turn-around time. The samples were transported following rigorous chain-of-custody guidelines to ensure proper handling and delivery of the samples. EMSL is accredited in the American Industrial Hygiene Association (AIHA) Environmental Microbiology Laboratory Accreditation Program (EMLAP) and is a successful participant in AIHA's Environmental Microbiology Proficiency Analytical Testing (EMPAT) program (Laboratory Number 102891.) The samples were analyzed via light microscopy at the standardized magnification of 600X. This technique does not allow for the differentiation between Aspergillus and Penicillium spores because they are morphologically identical. Additionally, the technique does not allow for cultivation, or the identification of spores to the species level, except in a few cases.

There are no universally accepted federal or State of Maryland standards for acceptable airborne concentrations of bioaerosols in an indoor occupational environment. In general, indoor airborne concentrations should be less than that found in the outdoor air, with similar species composition. Indoor spore counts significantly greater than those outdoors, or the presence of large numbers of different types of spores indoors that are not found outdoors, may indicate contamination and potential indoor air quality problems.

The total mold spore counts in all assessed areas of the school ranged between 710 spores/m³ and 51,670 spores/m³. The mold spore concentrations in the background sample obtained outdoors was 1,070 spores/m³. The total mold spore concentrations in the indoor samples obtained from Classroom 5 (sample # CLES-3), Classroom 14 (sample # CLES-6), Classroom 25 (sample # CLES-8) and the Multi-purpose room (sample # CLES-10) were between 1 and 2.5 times higher than the total mold spore concentration detected in the background sample (sample # CLES-BG.) Although the total mold spore concentrations in the above locations were higher than the total mold spore concentration in the background sample, the species composition of these samples were somewhat similar to the species composition of the background sample.

However, the total mold concentration in Classroom 17 (sample # CLES-7) was significantly (over 48 times) higher than the total mold spore concentration detected in the background sample. Furthermore, the concentration of *Aspergillus/ Penicillium* species detected in Classroom 17 (sample # CLES-7) was 51,500 spores/m³. No *Aspergillus/ Penicillium* spores were detected in the background sample (CLES-BG.) The significantly high concentration of *Aspergillus/ Penicillium* species detected in sample # CLES-7 indicates the presence of a potential indoor source(s) of mold in Classroom 17.



Aspergillus/ Penicillium are the most common mold species that are detected in indoor air samples. Most of the hundreds of sub-species are allergenic with only a few that are toxic. This group of species will grow with only the humidity in the air as its water source.

These areas were re-sampled on March 2, 2021 following cleanup activities. The results indicated that the total mold spore concentrations and the concentration of *Aspergillus/ Penicillium* spores in Classroom 17 and Classroom 25 were below the background concentration.

The summary of the results for the spore trap sampling are provided in Table 3 in **Attachment A**. The laboratory analytical results, including speciation and chain of custody forms for the spore trap samples are included in **Attachment B**.

CONCLUSIONS

- During the visual inspections conducted within typically occupied areas of the school, chosen by the Industrial hygienist's discretion, the follow issues were identified:
 - Classrooms 10, 5, 14, 17, 2 and Multipurpose Room: The ceiling-mounted air supply vents appeared to have dust accumulations and rust buildup.
 - Classroom 2: The supply grills of the window-mounted air conditioning unit appeared to have dust deposits. Accumulated dust was also observed on the air supply grills mounted on the wall of the classroom.
 - Classroom 14: The front panel of one of the window-mounted air conditioning unit was dismantled and appeared to be broken.
 - Health Unit: The window-mounted air conditioning unit was non-functional.
- Temperature levels recorded within all interior locations assessed, except in classroom 25, health unit and the multipurpose room were within the temperature levels typically observed during the fall-winter transitional period. The temperature levels in these locations were below the ASHRAE lower temperature standard of 68.0°F for winter months.
- The Relative humidity, CO₂, CO readings and particulate matter less than 10 microns (PM10) recorded within the assessed areas were within industry standards and guidelines;
- The total mold spore concentrations in all interior locations assessed were below the background sample concentration and were also consistent with those observed in the background sample. The results do not indicate elevated levels of airborne total fungal spores in the interior locations sampled.

RECOMMENDATIONS

Based on the results of our visual inspection, Tidewater proposes the following:

 Clean the ceiling-mounted air supply vents in Classrooms 10, 5, 14, 17, 2 and Multipurpose Room with a commercially available (EPA approved) disinfectant on a routine basis to remove dust deposits. If possible, also use a commercially available (EPA approved) rust remover to remove rust buildup from the supply vents;



- Clean the air supply grills of the window-mounted air conditioning unit and the air supply
 grills mounted on the walls of classroom 2 on a routine basis with a commercially available
 (EPA approved) disinfectant to remove dust deposits.
- The dismantled widow-mounted air-conditioning unit in Classroom 14 should be removed or replaced;
- The non-functional window-mounted air conditioning unit in the health unit should be repaired or replaced.
- Maintain good housekeeping practices in all common areas and classrooms. All common
 area and classrooms floors should be broom cleaned at the end of each day once the
 school re-opens for students. Furthermore, all horizontal surfaces including desktops,
 furniture, window sills, and light fixtures should be cleaned on a routine basis to prevent
 the accumulation of dust;
- Ensure the Heating Ventilation and Air Conditioning (HVAC) System supplying air to all common areas and classrooms is properly balanced per design requirements and are turned on and are operating at all times to ensure adequate ventilation throughout the classrooms before the school re-opens.

Qualifications

Tidewater has endeavored to investigate existing conditions in select areas of Cooper Lane Elementary School located at 3817 Cooper Lane in Landover Hills, Maryland as they pertain to indoor air quality and mold contamination. Our conclusions and recommendations are based on the observations made on the day of our assessment, laboratory data from the time of the assessment, and information provided by both our Client and the area occupants. Actual conditions vary from day to day throughout the year.

Tidewater appreciates the opportunity to provide Industrial Hygiene consulting services for Prince Georges County Public Schools. Please contact us should any questions arise concerning this report or if we may be of further assistance.

Sincerely,

Tidewater, Inc.

Skanda Abeyesekere, MS, CIH, CSP, CHMM

Skumber Arguverne

Project Manager

Jonathan N. Schatz, MS
Manager, IH Services

SA/JNS

Attachments: Attachment A – Summary of Comfort Parameters, PM10 Particulate Dust, and Microbial Results

Attachment B - Laboratory Reports and Chain of Custody Forms

Attachment C – Instrument Calibration Certificates

Attachment D - Relevant Certifications

Attachment E – Floor Plan with Sampling Locations



APPENDIX A

COMFORT PARAMETERS, PM10 PARTICULATE DUST, AND MICROBIAL RESULTS



Table 1: Indoor Air Quality Comfort Parameters Cooper Lane Elementary School									
Location	Temperature (°F)	Carbon Dioxide (ppm)	Relative Humidity (%)	Carbon Monoxide (ppm)					
	Novembe	r 19, 2020							
Main Office	68.2	29.3	578	0.4					
Library	71.5	28.4	546	0.4					
Classroom 10	72.5	21.3	501	0.3					
Classroom 5	73.1	23.1	564	0.2					
Classroom 14	70.1	24.0	498	0.0					
Classroom 17	69.7	25.4	506	0.1					
Classroom 25	57.8*	42.4	519	0.0					
Classroom 2	72.0	20.8	471	0.0					
Health Unit	67.8	26.7	476	0.0					
Multi-purpose Room	66.7	20.8	447	0.0					
Background (Outdoors)	52.1	28.9	440	0.0					

^{*}Highlighted Areas indicate locations in which temperature levels were below the American Society for Heating Refrigeration and Air Conditioning (ASHRAE) Standard 62.1 - 2019 recommended standards for winter months.



Table 2: Particulate Matter Less than 10 Microns (PM10) Cooper Lane Elementary School							
Location	Particulate Matter (PM10)						
Location	Concentration (mg/m³)						
November 19, 2020							
Main Office	0.083						
Library	0.071						
Classroom 10	0.073						
Classroom 5	0.074						
Classroom 14	0.074						
Classroom 17	0.074						
Classroom 25	0.071						
Classroom 2	0.080						
Health Unit	0.073						
Multipurpose Room	0.075						
Background (Outdoors)	0.074						



Table 3: Spore Trap Sampling Results Cooper Lane Elementary School

November 19, 2020

· · · · · · · · · · · · · · · · · · ·						
Sample Number	Sample Location	Sample Volume (L)	Total Fungi Concentration (Counts/m³)			
CLES-1	Main Office	75.0	830			
CLES -2	Library	75.0	990			
CLES-3	Classroom 5	75.0	1,240			
CLES-4	Classroom 10	75.0	850			
CLES-5	Classroom 2	75.0	710			
CLES-6	Classroom 14	75.0	1,160			
CLES-7	Classroom 17	75.0	51,670			
CLES-8	Classroom 25	75.0	2,300			
CLES-9	Health Unit	75.0	1,040			
CLES-10	Multipurpose Room	75.0	2,080			
CLES -BG	Background (Outdoors)	75.0	1,070			

^{*}Highlighted Area indicate location where the concentrations of the indoor sample exceeded the level detected in the background sample.



Table 3: Spore Trap Sampling Results Cooper Lane Elementary School March 2, 2021 Sample **Total Fungi** Volume **Sample Location** Concentration **Sample Number** (Counts/m³) (L) CLES-1 Classroom 25 75.0 40 CLES -2 Classroom 17 75.0 Noe Detected Background (Outdoors) CLES -BG 75.0 90



APPENDIX B LABORATORY REPORTS AND CHAIN OF CUSTODY FORMS



100 Green Park Industrial Court Saint Louis, MO 63123

Tel/Fax: (314) 577-0150 / (314) 776-3313

http://www.EMSL.com / saintlouislab@emsl.com

Attention: Skanda Abeyeskere

Tidewater, Inc. 6625 Selnick Drive

Suite A

Elkridge, MD 21075

EMSL Order: 392011029

Customer ID: TIDE50

Customer PO: Project ID:

Phone: (410) 540-8700

Fax: (410) 997-8713 **Collected Date:** 11/19/2020

Received Date: 11/30/2020 **Analyzed Date:** 12/07/2020

Project: Cooper Lane ES Test Report: Allergenco-D(™) Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391) Lab Sample Number 392011029-0001 392011029-0002 392011029-0003 Client Sample ID CLES-1 CLES-2 CLES-3 Volume (L) Sample Location: Main Office Library Classroom 5 **Spore Types Raw Count** Count/m³ % of Total **Raw Count** Count/m³ % of Total **Raw Count** Count/m³ % of Total Alternaria (Ulocladium) 2 90 10.8 5 200 20.2 3 100 8.1 Ascospores Aspergillus/Penicillium 2 90 9 1 40 32 1 440 15 660 66.7 8 300 Basidiospores 10 53 24.2 Bipolaris++ Chaetomium Cladosporium 8 300 36.1 40 18 790 63.7 Curvularia **Epicoccum** Fusarium Ganoderma Myxomycetes++ Pithomyces++ 1* 10* 8.0 Scopulariopsis/Microascus Stachybotrys/Memnoniella Unidentifiable Spores Zygomycetes Total Fungi 830 31 1240 20 100 23 990 100 100 Hyphal Fragment 1 40 10* Insect Fragment Pollen Analyt. Sensitivity 600x 44 44 44

13*

1

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

13*

1

1

Comber Steepmenn

No discernable field blank was submitted with this group of samples.

Amber Stegmann, Micro Supervisor or other Approved Signatory

13*

1

EMSL maintains liability limited to cost of analysis. Interpretation and use of test results are the responsibility of the client. This report relates only to the samples are proted above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling volumes and areas locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "." Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Saint Louis, MO

Analyt. Sensitivity 300x

Fibrous Particulate (1-4)

Skin Fragments (1-4)

Background (1-5)



100 Green Park Industrial Court Saint Louis, MO 63123

Tel/Fax: (314) 577-0150 / (314) 776-3313

http://www.EMSL.com / saintlouislab@emsl.com

Attention: Skanda Abeyeskere

Tidewater, Inc. 6625 Selnick Drive

Suite A

Elkridge, MD 21075 Project: Cooper Lane ES

EMSL Order: 392011029 Customer ID: TIDE50

Customer PO: Project ID:

Phone: (410) 540-8700 Fax: (410) 997-8713

Collected Date: 11/19/2020 **Received Date:** 11/30/2020

Analyzed Date: 12/07/2020

Test Report: Allergenco-D(™) Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)									
Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	392011029-0004 CLES-4 75 Classroom 10		3	392011029-0005 CLES-5 75 Classroom 2			392011029-0006 CLES-6 75 Classroom 14		
Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	2	90	10.6	-	-	-	-	-	-
Aspergillus/Penicillium	-	-	-	4	200	28.2	4	200	17.2
Basidiospores	15	660	77.6	5	200	28.2	10	440	37.9
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	3	100	11.8	7	300	42.3	10	440	37.9
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	1*	10*	1.4	1	40	3.4
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	1	40	3.4
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Total Fungi	20	850	100	17	710	100	26	1160	100
Hyphal Fragment	-	-	-	-	-	-	-	-	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	44	-	-	44	-	-	44	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	1	-	-	1	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific

Comber Stegmenn

No discernable field blank was submitted with this group of samples.

Amber Stegmann, Micro Supervisor or other Approved Signatory

EMSL maintains liability limited to cost of analysis, Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling

volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Saint Louis, MO



100 Green Park Industrial Court Saint Louis, MO 63123

Tel/Fax: (314) 577-0150 / (314) 776-3313

http://www.EMSL.com / saintlouislab@emsl.com

Attention: Skanda Abeyeskere

Tidewater, Inc. 6625 Selnick Drive

Suite A

Elkridge, MD 21075 Project: Cooper Lane ES

EMSL Order: 392011029 Customer ID: TIDE50

Customer PO: Project ID:

Phone: (410) 540-8700 Fax: (410) 997-8713

Collected Date: 11/19/2020 **Received Date:** 11/30/2020

Analyzed Date: 12/07/2020

Project: Coope	Cooper Lane ES								
Test Report: Aller	genco-D(™) Ana	alysis of Fungal	Spores & Part	ticulates by Opti	cal Microscopy	(Methods MIC	RO-SOP-201, A	STM D7391)	
Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	392011029-0007 CLES-7 75 Classroom 17			392011029-0008 CLES-8 75 Classroom 25			392011029-0009 CLES-9 75 Health Unit		
Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	-	-	-	-	-	-	1	40	3.8
Aspergillus/Penicillium	1180	51500	99.7	37	1600	69.6	3	100	9.6
Basidiospores	2	90	0.2	2	90	3.9	8	300	28.8
Bipolaris++	1	40	0.1	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	1	40	0.1	14	610	26.5	12	520	50
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	1	40	3.8
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	-	-	-	1	40	3.8
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Total Fungi	1184	51670	100	53	2300	100	26	1040	100
Hyphal Fragment	1	40	-	-	-	-	1	40	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	44	-	-	44	-	-	44	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	1	-	-	1	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific

Comber Stegmenn

Amber Stegmann, Micro Supervisor or other Approved Signatory

No discernable field blank was submitted with this group of samples.

EMSL maintains liability limited to cost of analysis. Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling

volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Saint Louis, MO



100 Green Park Industrial Court Saint Louis, MO 63123

Tel/Fax: (314) 577-0150 / (314) 776-3313

http://www.EMSL.com / saintlouislab@emsl.com

Attention: Skanda Abeyeskere

Tidewater, Inc. 6625 Selnick Drive

Suite A

Elkridge, MD 21075

Project: Cooper Lane ES

EMSL Order: 392011029 Customer ID: TIDE50

Customer ID: TIE

Project ID:

Phone: (410) 540-8700

Fax: (410) 997-8713

 Collected Date:
 11/19/2020

 Received Date:
 11/30/2020

 Analyzed Date:
 12/07/2020

Test Report: Aller Lab Sample Number: Client Sample ID:		92011029-0010 CLES-10	opoles a rail		92011029-0011 CLES-BG	(methods Mich	10-001-201, A	01.m D1001)	
Volume (L): Sample Location:		75 Multi Purpose			75 Outdoors				
Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	-	_	-
Alternaria (Ulocladium)	-	-	-	-	-	-		<u>'</u>	-
Ascospores	3	100	4.8	4	200	18.7			
Aspergillus/Penicillium	9	400	19.2	-	-	-			
Basidiospores	15	660	31.7	10	440	41.1			
Bipolaris++	-	-	-	-	-	-			
Chaetomium	-	-	-	-	-	-			
Cladosporium	19	830	39.9	6	300	28			
Curvularia	-	-	-	-	-	-			
Epicoccum	-	-	-	2	90	8.4			
Fusarium	-	-	-	-	-	-			
Ganoderma	-	-	-	-	-	-			
Myxomycetes++	2	90	4.3	1	40	3.7			
Pithomyces++	-	-	-	-	-	-			
Rust	-	-	-	-	-	-			
Scopulariopsis/Microascus	-	-	-	-	-	-			
Stachybotrys/Memnoniella	-	-	-	-	-	-			
Unidentifiable Spores	-	-	-	-	-	-			
Zygomycetes	-	-	-	-	-	-			
Total Fungi	48	2080	100	23	1070	100			
Hyphal Fragment	-	-	-	4	200	-			
Insect Fragment	-	-	-	-	-	-			
Pollen	-	-	-	-	-	-			-
Analyt. Sensitivity 600x	-	44	-	-	44	-			-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-			
Skin Fragments (1-4)	-	1	-	-	1	-			
Fibrous Particulate (1-4)	-	1	-	-	1	-			
Background (1-5)	-	1	-	-	1	-			

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

amber Stegmenn

No discernable field blank was submitted with this group of samples.

Amber Stegmann, Micro Supervisor or other Approved Signatory

EMSL maintains liability limited to cost of analysis. Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling volumes and areas. locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification.

Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "" Denotes particles found at 300X." Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Saint Louis, MO

Microbiology Chain of Custody EMSL Order Number (Lab Use Only):

		39201	1 029			PHONE: FAX:
Company: Tidew	ater Inc			EM If Bill to		ifferent Same ructions in Comments**
Street: 6625 Selnick	Drive, Suite A			Third Party Bill	ing requires written	authorization from third party
City: Elkridge	St	ate/Province:	MD Zi	p/Postal Code	9;	Country:
	Skanda Abeyesekere	,	Te	lephone #:		
	anda@tideh2o.net			x #:	Pt	urchase Order:
Project Name/Number		e ES		ease Provide	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	AX E-mail Mail
U.S. State Samples T						nercial 🔲 Residential
	Turna	round Time (TAT) Options*	- Please Che	ck	
	6 Hour 🔳 24 Hour	☐ 48 Hot	ır 🔲 72 H	our 🔲 96	Hour 1	Week 2 Week
*Analysis completed in a						ect to methodology requirements
			ples (Spore			T 11470 \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \
M001 Air-O-Cell M049 BioSiS	 M173 Allegro M2 M003 Burkard 	• M004 /	Allergenco	M032 AllM002 Cy		M172 Versa Trap
• M030 Micro 5	M174 MoldSnap		Relle Smart	• M130 Via		11
		Other Mici	robiology Tes	t Codes		
 M041 Fungal Direc 			Indotoxin Analy			nterpcocci
.M005 Viable Fungi M006 Viable Fungi			leterotrophic Pl Real Time Q-PC			cal Coliform
M007 Culturable Fungi	ID and Count (Speciation)	M180 Panel	Real Time Q-PC	K-EKIVII 30		RSA Analysis yptpcoccus neoformans
M008 Culturable Fu			otal Coliform		Detection	n
M009 Gram Stain C			Membrane Filtr			stoplasma capsulatum
INU10 Bacterial Col Prominent	unt and ID – 3 Most		ecal <i>Streptocol</i> Membrane Filtr		Detection M033-39	Allergen Testing
M011 Bacterial Co.	unt and ID 5 Most		215 Legionella I			oup Allergen
Prominent			Recreational Wa			og, Cockroach, Dustmites)
	tamination in Buildings	• M027 N	/lycotoxin Analy	sis	Other Se	ee Analytical Price Guide
Preservation Method	(Water):					
Sk	anda Abeyesekere		Ka	elle A	1000	
Name of Sampler:				ure of Sample		
Sample #	Sample Location	_	Sample Type	Test Code	Volume/Area	
Example: A1				M001	75L 3 3 1 1	1/1/12 4:00 PM
CLES-1	Main Office	<u></u>	Arr	m632	75-0	11/19/2020
CLES-2	hibrary		1			
CLES-3	Classoon	<u>ح</u>		└		
C4ES-4	ClassRoom	10				_
CUES-15	C (C 25 1000	2		<u> </u>		<u> </u>
LLES-6	C10251000	10		,	·	
CLE3-7	CKSSroom	17				
		<u> </u>				TE Just
			4		4	
Client Sample # (s):	11		То	tal # of Sampl	les: //	NOV EAR
Relinquished (Client)		le	Date: //	/19/202	O Time: (Q.	SVILLY SVILLY
Received (Client):	. Janvarth Lee	17x	Date:	1	Time:	IVE LE,
Comments:	- June June		- ate.		111161	<u> </u>
•	•					. 03 NC
						6/9

OrderID: 392011029

Microbiology Chain of Custody EMSL Order Number (Lab Use Only):

		
1	200 - 11 000 1	
	702011029 I	

PHONE: FAX:

Additional Pages of the Chain of Custody are only necessary if needed for additional sample information

Sample #	Sample Location	Sample Type	Test Code	Volume/Area	Date/Time Collected
-LES-8	elassons 25 Heath Unit Multi: furpose ouldoss	Acc			
L€S-9	Heath unit				
LES-10	Multi Purpose				
LES-BY	outdoors	1	, .		
		<u> </u>			
			ļ		·
		<u> </u>			
			·		
,		ļ: <u> </u>			·
		<u> </u>		·	
		ļ			
	•				
1					<u> </u>
	·				
·		ļ			·
<u> </u>					
	· 		· ·		
					1
1 -		-			· .
		-			
		-			
omments/Special	Instructions:				¥
ommonw/opeolal	mod adultion				
i					

Page 2 of 2 pages



10768 Baltimore Avenue Beltsville, MD 20705 Tel/Fax: (301) 937-5700 / (301) 937-5701

http://www.EMSL.com / beltsvillelab@emsl.com

Attention: Skanda Abeyeskere

Tidewater, Inc. 6625 Selnick Drive

Suite A

Elkridge, MD 21075

Project: PGCPS COOPER LANE ES

EMSL Order: 192101966

Customer ID: TIDE50 **Customer PO:**

(410) 997-8713

Project ID:

(410) 540-8700

03/02/2021 Collected Date: Received Date: 03/02/2021 Analyzed Date: 03/05/2021

Test Report: Allergenco-D(™) Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391) Lab Sample Number 192101966-0001 192101966-0002 192101966-0003 Client Sample ID CLES-1 CLES-1 **CLES-BG** Volume (L) 75 75 75 Sample Location **CLASSRM 25 CLASSRM 17 OUTDOORS Spore Types Raw Count** Count/m³ % of Total **Raw Count** Count/m³ % of Total **Raw Count** Count/m³ % of Total Alternaria (Ulocladium) 40 100 Ascospores 1 Aspergillus/Penicillium 2 90 Basidiospores 100 Bipolaris++ Chaetomium Cladosporium Curvularia **Epicoccum** Fusarium Ganoderma Myxomycetes++ Pithomyces++ Scopulariopsis/Microascus Stachybotrys/Memnoniella Unidentifiable Spores Zvgomvcetes Total Fungi 40 100 90 100 Hyphal Fragment 1 40 Insect Fragment Analyt. Sensitivity 600x 44 44 44 Analyt. Sensitivity 300x 13 13* 13*

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific

2

1

No discernable field blank was submitted with this group of samples.

Skin Fragments (1-4) Fibrous Particulate (1-4)

Background (1-5)

1

2

Abubakar Barry, Microbiology Lab Manager or other Approved Signatory

EMSL maintains liability limited to cost of analysis, Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling

volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted.

High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

1

Samples analyzed by EMSL Analytical, Inc. Beltsville, MD AIHA-LAP, LLC-EMLAP Accredited #102891

Initial report from: 03/05/2021 02:29 PM

Microbiology Chain of Custody EMSL Order Number (Mach Use Only):

192101966	

PHONE: FAX:

Company.	ater Inc.				SL-Bill to: Dif	ferent Same ions in Comments**
Street: 6625 Selnick	Drive, Suite A			Third Party Billi	ing requires written au	thorization from third party
City: Elkridge		ate/Province:	MD z	ip/Postal Code	- $ -$	ountry:
report to (Manie).	kanda Abeyesekere		Τ,	elephone#:		·
Email Address: Ska	anda@tideh2o.net		F	ax #:	Purc	hase Order:
Project Name/Numbe	r: PGCPS COOPES	Lane	ES P	lease Provide	Results: FAX	E-mail Mail
U.S. State Samples T	1		- I	onnecticut Sa	mples: 🗌 Comme	rcial Residential
	. Turna	round Time (TATL Options	* - Please Che	ck	
<u></u>	6 Hour 🔳 24 Hour	☐ 48 Hou			Hour 1 V	
*Analysis completed in ac	cordance with EMSL's Terms					to methodology requirements
				Traps) - Tes		
 M001 Air-O-Cell M049 BioSIS 	 M173 Allegro M2 M003 Burkard 	• MO43 C	Allergénco Svoley	 M032 Alle M002 Cy 		M172 Versa Trap
• M030 Micro 5	M174 MoldSnap		Relie Smart	■ M130 Via		, <u> </u>
	-,		obiology Te	st Codes		
M041 Fungal Direct	Examination		ndotoxin Anal		• M029 Ente	rococci
M005 Viable Fungi			leterotrophic F		 M019 Feca 	
_=	ID and Count (Speciation)		Real Time Q-P	CR-ERMI 36	• M133 MRS	
 M007 Culturable Fu M008 Culturable Fu 		 Panel M018 T 	otal Coliform		M028 Cryp Detection	tococcus neoformans
M009 Gram Stain C		****	Membrane Fill	ration)	_ ,	plasma capsulatum
M010 Bacterial Cou	int and ID – 3 Most	• M020 F	ecal Streptoco	occus .	Detection	
Prominent			Membrane Fili			llergen Testing
M011 Bacterial Cou Prominent	int and ID - 5 Most		1 15 <i>Legionella</i> Recreational W		M044 Grou (Cat Dog	Cockroach, Dustmites)
M013 Sewage Cont	tamination in Buildings		iycotoxin Ana			Analytical Price Guide
· · · · · · · · · · · · · · · · · ·						
I Preservation Method	(Water):		,			-
Preservation Method				Is-l	2 4 - 2	
SLAMDA	(Water): ASEYESE	16 V26	Signa	ture of Sample	hy hy	
SLOW) Name of Sampler:	ASEYESE		Sample	ture of Sample	er; John Volume/Area	Date/Time Collected
S LOWDA Name of Sampler: Sample #	ASEYESE Sample Location	on	Sample Type	Test Code	Volum <i>el</i> Area	
SLOWDO Name of Sampler: Sample #	Sample Location	on	Sample Type	Test Code M001	Volume/Area	1/1/1/2:4:00 PM
S LOWDA Name of Sampler: Sample #	ASEYESE Sample Location	on	Sample Type	Test Code	Volum <i>el</i> Area	
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1	Sample Location Kitchen Classnoom	on	Sample Type Air	Test Code M001 M032	75L 7-5	1/1/1/2:4:00 PM
SLOWDO Name of Sampler: Sample # Example: At	Sample Location	on	Sample Type	Test Code M001	Volume/Area	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: At CLES-1 CLES-2	Sample Location Kitchen Classmon Classmon	on 25	Sample Type Air Air	Test Code M001 M032	Volume/Area 75L 7-5 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1	Sample Location Kitchen Classmon Classmon	on 25	Sample Type Air Ar	Test Code M001 M032	75L 7-5	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2	Sample Location Kitchen Classmon Classmon	on 25	Sample Type Air Air	Test Code M001 M032	Volume/Area 75L 7-5 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2	Sample Location Kitchen Classmon Classmon	on 25	Sample Type Air Air	Test Code M001 M032	Volume/Area 75L 7-5 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2	Sample Location Kitchen Classmon Classmon	on 25	Sample Type Air Air	Test Code M001 M032	Volume/Area 75L 7-5 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2 CLES-B6	Sample Location Kitchen Classnoon Classnoon Dutdows -	on 25	Sample Type Air Air A	Test Code M001 M032 M032 M032	Volume/Area 751 75 75 75	1/1/1/2 4:00 PM
SLAMDA Name of Sampler: Sample # Example: At CLES-1 CLES-2	Sample Location Kitchen Classnoon Classnoon Dutdows -	on 25	Sample Type Air Air A	Test Code M001 M032	Volume/Area 751 75 75 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2 CLES-BG Client Sample # (s):	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25	Sample Type Air Arr	Test Code M001 M032 M032 M032 M032 M032	Volume/Area 75L 7-5 75 75	1/1/1/2:4:00 PM
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2 CLES-BG Client Sample # (s): Relinquished (Client)	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25	Sample Type Air Air Air Date: C	Test Code M001 M032 M032 M032	Volume/Area 751 75 75 75 75 75 75 75 75 7	1/1/1/2 4:00 PM 03 02 2:02
Name of Sampler: Sample # Example: A1 CLES-1 CLES-BG Client Sample # (s): Relinquished (Client):	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25	Sample Type Air Arr	Test Code M001 M032 M032 M032 M032 M032	Volume/Area 75L 7-5 75 75	1300 PM 03 102 1202
SLAMDA Name of Sampler: Sample # Example: A1 CLES-1 CLES-2 CLES-BG Client Sample # (s): Relinquished (Client)	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25	Sample Type Air Air Air Date: C	Test Code M001 M032 M032 M032 M032 M032	Volume/Area 751 75 75 75 75 75 75 75 75 7	1300 By W.
Name of Sampler: Sample # Example: At CLES-1 CLES-BG Client Sample # (s): Relinquished (Client):	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25	Sample Type Air Air Air Date: C	Test Code M001 M032 M032 M032 M032 M032	Volume/Area 751 75 75 75 75 75 75 75 75 7	111/1/2/400 PM RICEIVED RISK ANALYTINAL BELT WILLE, IV
Name of Sampler: Sample # Example: At CLES-1 CLES-BG Client Sample # (s): Relinquished (Client):	Sample Location Kitchen Classnoon Classnoon Dutdows 3	on 25 17 Background	Sample Type Air Air Air Date: C	Test Code M001 M032 M032 M032 M032 M032 M032 M032	Volume/Area 751 75 75 75 75 75 75 75 75 7	300 EU N



APPENDIX C INSTRUMENT CALIBRATION CERTIFICATES



CERTIFICATE OF CALIBRATION AND TESTING

TSI Incorporated, 500 Cardigan Road, Shoreview, MN 55126 USA Tel: 1-800-874-2811 1-651-490-2811 Fax: 1-651-490-3824 http://www.tsi.com

ENVIRONMENT CONDITIONS			Money	OFCE V	
TEMPERATURE	74.1 (23 4)	°F (°C)	MODEL	9565-X	
RELATIVE HUMIDITY	26	%RH		050574045000	
BAROMETRIC PRESSURE	29.26 (990.9)	inHg (nPa)	SERIAL NUMBER	9565X1945002	

- CALIBRATION VERIFICATION RESULTS-

THERMO COUPLE^			SYSTE	Unit: °F (°C)			
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	71.6 (22.0)	71.6 (22.0)	69.6~73.6 (20.9~23.1)				

BAROMETRIC PRESSURE		ESSURE	SYSTEM PRESSURE01-01				Unit: inHg (hPa)	
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE	
1	29.26 (990.9)	29.26 (990.9)	28.67~29.85 (970.9~1010.8)					

[^] Circuit portion of temperature measurement only, not including probe.

TSI does hereby certify that the above described instrument conforms to the original manufacturer's specification (not applicable to As Found data) and has been catibrated using standards whose accuracies are traceable to the United States National Institute of Standards and Technology (NIST) or has been verified with respect to instrumentation whose accuracy is traceable to NIST, or is derived from accepted values of physical constants. TSI's calibration system is registered to ISO-9001:2615

Measurement Variable	System ID	Last Cal.	Cal. Due	Measurement Variable	System ID	Last Cal.	Cal. Due
DC Voltage	E003299	06-06-19	12-31-20	DC Voltage	E003300	06-06-19	12-31-20
Temperature	E004626	01-09-19	01-31-20	Pressure	E003302	08-07-19	02-29-20
Pressure	E003303	08-26-19	02-29-20				

Rose Germain

November 8, 2019

DATE

DOC. ID. CERT_GEN_WCC_TM



CERTIFICATE OF CALIBRATION AND TESTING

TSI Incorporated, 500 Cardigan Road, Shoreview, MN 55126 USA Tel: 1-800-874-2811 1-651-490-2811 Fax: 1-651-490-3824 http://www.tsi.com

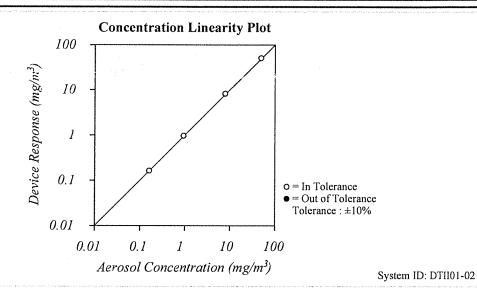
Environment Conditions		
Temperature	75.83 (24.4)	°F (°C)
Relative Humidity	43.6	%RH
Barometric Pressure	28.93 (979.7)	inHg (hPa)

Model	8534
Serial Number	8534170101

 ☑ As Left
 ☑ In Tolerance

 ☐ As Found
 ☐ Out of Tolerance





FLOW AND PRESSURE VERIFICATION SYSTEM DTII01-01 Measured **Parameter** Standard Allowable Range Parameter Standard Measured Allowable Range Flow lpm 3.00 3.03 2.88 ~ 3.12 Pressure kPa 97.8 97.8 92.95 ~ 102.73 Full Flow Ipm N/A 4.54 >3.80

TSI Incorporated does hereby certify that all materials, components, and workmanship used in the manufacture of this equipment are in strict accordance with the applicable specifications agreed upon by TSI and the customer and with all published specifications. All performance and acceptance tests required under this contract were successfully conducted according to required specifications. There is no NIST standard for optical mass measurements. Calibration of this instrument performed by TSI has been done using emery oil and has been nominally adjusted to respirable mass per standard ISO 12105-1, At test dust (Arizona dust). Our calibration ratio is greater than 1.2:1

Measurement Variable	System ID	Last Cal.	Cal. Due
DC Voltage	E003314	01-15-20	01-31-21
Microbalance	M001324	10-03-18	10-31-20
3 um PSL	221853	n/a	n/a
Pressure	E003511	10-04-19	10-31-20
DC Voltage	E003315	01-15-20	01-31-21
Flowmeter	E005922	06-29-20	06-30-21
Microbalance	M001324	10-03-18	10-31-20
1 um PSL	698880	n/a	n/a
10 um PSL	212455	n/a	n/a

	01 00 00	^1 ^1 ^1
Pressure E005651	01-09-20 09-15-20 06-15-20 07-06-20 n/a	01-31-21 03-31-21 06-30-21 07-31-21 n/a

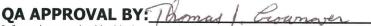
David Farrell

September 24, 2020

Date



The instrument listed above is in conformance with factory specifications and the flow is set to nominal using a BUCK Calibrator which is N.I.S.T. traceable to A. P. Buck, Inc. Calibration Procedure APB-1, Ver. 6.2.



Information contained in this document should not be reproduced in any form without the written consent of A.P. Buck Inc. It is for reference only and cannot be used as a form of endorsement by any private or governmental regulatory body.

A.P. BUCK, INC.
7101 Presidents Drive, Suite 110
Orlando, FL 32809
Phone: 407-851-8602
• Fax: 407-851-8910



COCR-004 REV-01 3/3/2006























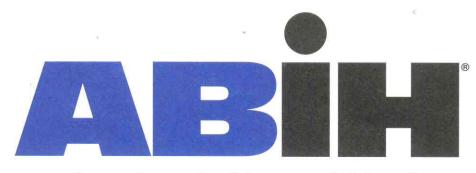








APPENDIX D RELEVANT CERTIFICATIONS



american board of industrial hygiene®

organized to improve the practice of industrial hygiene proclaims that

Skandakumar Harshanath Abeyesekere

having met all requirements of education, experience and examination, and ongoing maintenance, is hereby certified in the

of INDUSTRIAL HYGIENE

and has the right to use the designations

CERTIFIED INDUSTRIAL HYGIENIST

CIH

Certificate Number

9928 CP

Awarded:

May 11, 2011

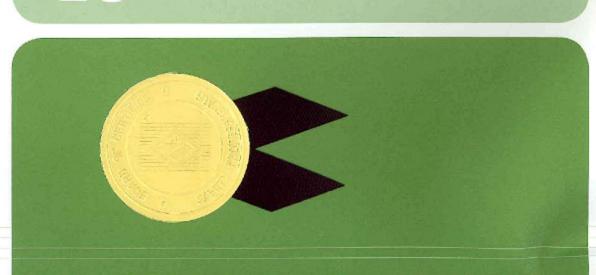
Expiration Date:

December 1, 2021

A 3- 13-

Chair, ABIH

Chief Executive Officer, ABIH



CERTIFIED SAFETY PROFESSIONALS **BOARD OF**

affirms that

Skandakumar Abeyesekere

Has applied for, met qualifications, and passed required examination(s) and is hereby authorized to use the designation

Certified Safety Professional®

in Comprehensive Practice

So long as this certificate is not suspended or revoked and the certificant renews this authorization annually and meets Continuance of Certification requirements. Board of Examiners in witness whereof we have here unto set our hands and affixed the Seal of the Board this 7th Day of April, 2008



President

Secretary

20110

CSP No.

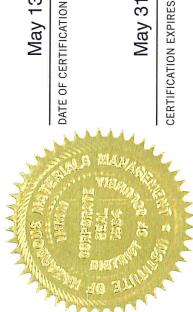


THIS CERTIFIES THAT

Skandakumar Abeyeskere

HAS SUCCESSFULLY MET ALL THE REQUIREMENTS OF EDUCATION, EXPERIENCE AND EXAMINATION, AND IS HEREBY DESIGNATED A

CERTIFIED HAZARDOUS MATERIALS MANAGER C E C E



May 13, 2016

DATE OF CERTIFICATION

May 31, 2021

CREDENTIAL NUMBER

M. Patricia Buly

ACTING EXECUTIVE DIRECTOR



Accredited by the American National Standards Institute and the Council of Engineering and Scientific Specialty Boards





APPENDIX E

FLOOR PLAN WITH SAMPLING LOCATIONS

