

January 11, 2021

Prince George's County Public Schools
Environmental Safety Office
13306 Old Marlboro Pike
Upper Marlboro, MD 20772

Attention: Alex Baylor
alex.baylor@pgcps.org

Subject: Indoor Air Quality Survey
John Eager Howard Community Center
4400 Shell Street
Capitol Heights, MD 20743

Mr. Baylor:

On December 4, 2020, a Soil and Land Use Technology, Inc. (SaLUT) Industrial Hygienist conducted an indoor air quality (IAQ) evaluation at John Eager Howard Community Center, a property maintained by Prince George's County Public Schools (PGCPS) located at 4400 Shell Street, Capitol Heights, MD 20743. The inspection was performed in accordance with PGCPS contract number IFB 022-19.

Methodology

The IAQ evaluation conducted by SaLUT included a visual assessment, IAQ instrumentation screening, and a collection of interior air samples for mold in representative locations throughout the building. Additionally, one building exterior environmental air sample was taken for comparison.

Air-borne fungal spore samples were collected on *Air-O-Cell* cassettes using a Buck BioAire calibrated pump. The air samples were taken between three and five feet from the ground. In tandem with collecting mold samples, real-time readings for carbon dioxide, carbon monoxide, temperature and relative humidity were collected using a Fluke 975 Air Meter in representative areas within the facility.

The fungal spore air samples were delivered to EMSL Analytical, Inc. of Beltsville, Maryland for analysis. Fungal spores and particulates in air samples were analyzed by Optical Microscopy (methods EMSL 05-TP-003 and ASTM D7391). The sample chain-of-custody and laboratory reports are attached.

Observations

The table below summarizes the main observations from the IAQ survey at John Eager Howard Community Center, visited on December 4, 2020.

Table 1-Observations

Location	Summary of Observations 12-4-2020
Cafeteria	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth; Mild odor; Stained ceiling tiles; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
Hallway Between Classroom 1 and 11	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
Hallway Between Classroom 05 and 06	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
In front of Classroom 06	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
Conference and Planning Room	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.

Measurements of Indoor Environmental Quality Parameters

Table 2 depicts a summary of average measurements of comfort.

Temperature

The American Society of Heating, Refrigerating and Air-Conditioning Engineers (ASHRAE) have published recommendations for year round acceptable temperatures in Standard 55-2010 *Thermal Environmental Conditions for Human Occupancy*. The winter comfort range is 20 to 24°C (68 to 75°F) and 23 to 26°C (73 to 79°F) is the summer comfort range. The temperature readings were within the ASHRAE recommended ranges in the representative spaces with the exception of the Hallway Between Classroom 1 and 11.

Relative Humidity (RH)

RH is a key factor for mold growth. Mold has the potential of growing on suitable surfaces with humidity levels above 60%. ASHRAE Standard 62.1-2010 *Ventilation for Acceptable Indoor Air Quality* recommends a maximum indoor RH of 65% to preclude the likelihood of condensation on cool surfaces encouraging mold growth. The RH readings were within the ASHRAE recommended ranges in the representative areas.

Carbon Dioxide (CO₂)

Under conditions of maximum occupancy, ASHRAE Standard 62.1-2010, Appendix C, infers that the acceptable CO₂ upper limit is the prevailing outdoor CO₂ concentration plus 700 parts per million (ppm). On the day of the space evaluation, the outdoor (building exterior) CO₂ concentration was approximately 469 ppm therefore indoor concentrations should not exceed approximately 1,169 ppm (700 + 469). The maximum average interior CO₂ concentration detected was 645 ppm in front of Classroom 06, a range within the ASHRAE recommendations, per Table 2 below.

Carbon Monoxide (CO)

CO is a colorless and odorless gas that is produced by the incomplete combustion of carbon containing fuels. Oil, gasoline, diesel fuels, wood, coke, and coal are major sources of CO. All registered CO concentrations were below the EPA National Ambient Air Quality Standard (NAAQS) of 9 ppm, per Table 2 below.

**Table 2: John Eager Howard Community Center, Instrumental Screening Levels
December 4, 2020 (7:30 AM-9:30 AM)**

Sample Location	Temp °F	RH%	CO ppm	CO ₂ ppm
Standards	ASHRAE 68 to 75°F*	ASHRAE <65%	NAAQS 9	ASHRAE 1,169
Cafeteria	68.0	29.9	0	538
Hallway Between Classroom 1 and 11	66.9	27.7	0	490
Hallway Between Classroom 05 and 06	72.4	25.2	0	505
In front of Classroom 06	69.0	24.3	0	645
Conference and Planning Room	71.3	23.5	0	493
Outside Exterior EV Sample	60.8	39.0	0	469

PM - Particulate Matter size
°F - Degrees Fahrenheit
CO - Carbon Monoxide
ppm - parts per million

µg/m³ - micrograms per cubic meter
RH% - % Relative Humidity
CO₂ - Carbon Dioxide
* - Winter Comfort Range

Mold-in-Air Samples

There are no definitive regulations or standardized guidelines for addressing airborne mold in an indoor setting. If building systems (ventilation, envelope) are functioning properly, the indoor population profile should mimic what is encountered outdoors and the concentrations should be below the outdoor (building exterior) environmental sample levels.

Table 3 summarizes airborne mold spore sampling results and locations. On December 4, 2020, total mold counts in representative samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations. Laboratory analysis follows this report (see attachment).

**Table 3: John Eager Howard Community Center -
Measurements of Mold-in-Air Samples
December 4, 2020 (7:30 AM-9:30 AM)**

Spore Types	Cafeteria	Hallway between Classrooms 1 and 11	Hallway between Classroom 05 and 06	In front of Classroom 06
<i>Alternaria (Ulocladium)</i>	-	-	-	-
<i>Ascospores</i>	40	-	-	-
<i>Aspergillus/Penicillium</i>	40	200	40	100
<i>Basidiospores</i>	400	300	200	590
<i>Bipolaris++</i>	-	-	-	-
<i>Chaetomium</i>	-	-	-	-
<i>Cladosporium</i>	80	-	-	-
<i>Curvularia</i>	-	-	-	-
<i>Epicoccum</i>	-	-	-	-
<i>Fusarium</i>	-	-	-	-
<i>Ganoderma</i>	-	-	-	-
<i>Myxomycetes++</i>	-	10	-	-
<i>Pithomyces++</i>	-	-	-	-
<i>Rust</i>	-	-	-	-
<i>Scopulariopsis/Microascus</i>	-	-	-	-
<i>Stachybotrys/Memnoniella</i>	-	-	-	-
<i>Unidentifiable Spores</i>	-	-	-	-
<i>Zygomycetes</i>	-	-	-	-
<i>Nigrospora</i>	-	-	-	-
<i>Hyphal Fragment</i>	-	-	-	-
<i>Insect Fragment</i>	40	-	-	-
<i>Pollen</i>	-	-	-	-
Total Fungi	560	510	240	690

* Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

**Table 3: John Eager Howard Community Center -
Measurements of Mold-in-Air Samples continued
December 4 2020 (7:30 AM-9:30 AM)**

Spore Types	Conference and Planning Room	Outside Exterior EV Sample	Field Blank
<i>Alternaria (Ulocladium)</i>	40	40	-
<i>Ascospores</i>	-	80	-
<i>Aspergillus/Penicillium</i>	-	200	-
<i>Basidiospores</i>	40	2,300	-
<i>Bipolaris++</i>	-	-	-
<i>Chaetomium</i>	-	-	-
<i>Cladosporium</i>	10	1,900	-
<i>Curvularia</i>	-	40	-
<i>Epicoccum</i>	-	100	-
<i>Fusarium</i>	-	-	-
<i>Ganoderma</i>	-	-	-
<i>Myxomycetes++</i>	-	300	-
<i>Pithomyces++</i>	-	-	-
<i>Rust</i>	-	930	-
<i>Scopulariopsis/Microascus</i>	-	-	-
<i>Stachybotrys/Memnoniella</i>	-	-	-
<i>Unidentifiable Spores</i>	-	-	-
<i>Zygomycetes</i>	-	-	-
<i>Nigrospora</i>	-	-	-
<i>Hyphal Fragment</i>	-	40	-
<i>Insect Fragment</i>	-	-	-
<i>Pollen</i>	-	-	-
Total Fungi	90	7,830	No Trace

*Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

Findings and Conclusions

The comfort parameters (i.e., temperature, RH, CO₂, and CO levels) in the representative areas conform to ASHRAE and/or NAAQS guidelines. On December 4, 2020, total mold counts in representative area samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations, indicating no amplified mold growth.

Thank you for the opportunity to provide industrial hygiene services for PGCPs. If you have any questions, please contact me at 301.595.3783.

Sincerely,



Chaminda Jayatilake, PE, CIH, CSP, CHMM
Certified Industrial Hygienist
Soil and Land Use Technology Inc. (SaLUT)

Attachment

Attachment - Mold Spore Sample Analytical Results and Chain-of-Custody Forms

Attachment

Mold Spore Sample Analytical Results and Chain-of-Custody Forms



EMSL Analytical, Inc.

5221 Militia Hill Road Plymouth Meeting, PA 19462
Tel/Fax: (610) 828-3102 / (610) 828-3122
<http://www.EMSL.com> / plymouthmeetinglab@emsl.com

EMSL Order: 182003920
Customer ID: SALU50
Customer PO:
Project ID:

Attention: Indika Jayatilake
SaLUT
1818 New York Avenue, NE
Suite 231
Washington, DC 20002
Project: 19-035 John Edger Howard

Phone: (301) 595-3783
Fax: (301) 595-3787
Collected Date: 12/04/2020
Received Date: 12/04/2020 01:06 PM
Analyzed Date: 12/09/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	182003920-0001 S1 75 Cafeteria			182003920-0002 S2 75 HW Between CR 11 and 12			182003920-0003 S3 75 Infront of CR 6A			
	Spore Types	Raw Count	Count/M³	% of Total	Raw Count	Count/M³	% of Total	Raw Count	Count/M³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-	-
Ascospores	1	40	7.1	-	-	-	-	-	-	-
Aspergillus/Penicillium	1	40	7.1	4	200	39.2	3	100	14.5	
Basidiospores	9	400	71.4	6	300	58.8	14	590	85.5	
Bipolaris++	-	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-	-
Cladosporium	2	80	14.3	-	-	-	-	-	-	-
Curvularia	-	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	1*	10*	2	-	-	-	-
Pithomyces++	-	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-	-
Arthrinium	-	-	-	-	-	-	-	-	-	-
Oidium	-	-	-	-	-	-	-	-	-	-
Total Fungi	13	560	100	11	510	100	17	690	100	
Hyphal Fragment	-	-	-	-	-	-	-	-	-	-
Insect Fragment	1	40	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-	-
Skin Fragments (1-4)	-	2	-	-	2	-	-	1	-	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-	-
Background (1-5)	-	1	-	-	1	-	-	1	-	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Kevin Ream, Laboratory Manager
or other Approved Signatory

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Samples analyzed by EMSL Analytical, Inc. Plymouth Meeting, PA AIHA-LAP, LLC-EMLAP Accredited #178659

Initial report from: 12/09/2020 12:10 PM

For information on the fungi listed in this report, please visit the Resources section at www.emsl.com



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Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	182003920-0004			182003920-0005			182003920-0006		
	S4	S5	S6	Room Conference and Planning			Outside		
	75	75	75						
	HW Between CR 05 and 06			Room Conference and Planning			Outside		
Spore Types	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total
Alternaria (Ulocladium)	-	-	-	1	40	44.4	1	40	0.5
Ascospores	-	-	-	-	-	-	2	80	1
Aspergillus/Penicillium	1	40	16.7	-	-	-	5	200	2.6
Basidiospores	5	200	83.3	1	40	44.4	54	2300	29.4
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	-	-	-	1*	10*	11.1	44	1900	24.3
Curvularia	-	-	-	-	-	-	1	40	0.5
Epicoccum	-	-	-	-	-	-	3	100	1.3
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	-	-	-	6	300	3.8
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	22	930	11.9
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Arthriniium	-	-	-	-	-	-	1	40	0.5
Oidium	-	-	-	-	-	-	46	1900	24.3
Total Fungi	6	240	100	3	90	100	185	7830	100
Hyphal Fragment	-	-	-	-	-	-	1	40	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	2	-	-	1	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Kevin Ream, Laboratory Manager
or other Approved Signatory

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Samples analyzed by EMSL Analytical, Inc. Plymouth Meeting, PA AIHA-LAP, LLC-EMLAP Accredited #178659

Initial report from: 12/09/2020 12:10 PM

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Project: 19-035 John Edger Howard

Phone: (301) 595-3783
Fax: (301) 595-3787
Collected Date: 12/04/2020
Received Date: 12/04/2020 01:06 PM
Analyzed Date: 12/09/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	182003920-0007		
Client Sample ID:	S7		
Volume (L):			
Sample Location:	Field Blank		
Spore Types	Raw Count	Count/M³	% of Total
Alternaria (Ulocladium)	-	-	-
Ascospores	-	-	-
Aspergillus/Penicillium	-	-	-
Basidiospores	-	-	-
Bipolaris++	-	-	-
Chaetomium	-	-	-
Cladosporium	-	-	-
Curvularia	-	-	-
Epicoccum	-	-	-
Fusarium	-	-	-
Ganoderma	-	-	-
Myxomycetes++	-	-	-
Pithomyces++	-	-	-
Rust	-	-	-
Scopulariopsis/Microascus	-	-	-
Stachybotrys/Memnoniella	-	-	-
Unidentifiable Spores	-	-	-
Zygomycetes	-	-	-
Arthriniium	-	-	-
Oidium	-	-	-
Total Fungi	-	No Trace	-
Hyphal Fragment	-	-	-
Insect Fragment	-	-	-
Pollen	-	-	-
Analyt. Sensitivity 600x	-	0	-
Analyt. Sensitivity 300x	-	0*	-
Skin Fragments (1-4)	-	-	-
Fibrous Particulate (1-4)	-	-	-
Background (1-5)	-	-	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Kevin Ream, Laboratory Manager
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Microbiology Chain of Custody

EMSL Order Number (Lab Use Only):

182003920

EMSL ANALYTICAL, INC.
 200 ROUTE 130 NORTH
 CINNAMINSON, NJ 08077
 PHONE: (800) 220-3675
 FAX: (856) 786-0262



EMSL ANALYTICAL, INC.
 LABORATORY PRODUCTS TRAINING

Company Name: Salut Inc
Street: 1818 New York Ave NE Suite 231
City: Washington **State/Province:** DC **Zip/Postal Code:** **Country:**
Report To (Name): Indira Jayatilake **Telephone #:**
Email Address: ijayatilake@yahoo.com **Fax #:** **Purchase Order:**
Project Name/Number: 14-035 - John Edger Howard **Please Provide Results:** Fax Email
U.S. State Samples Taken: MD **Project Zip Code:** 20743 **Connecticut Samples:** Commercial Residential
Sterile, Sodium Thiosulfate Preserved Bottle Used: **Biocide Used in Source (specify):**
Public Water Supply Samples: **Note:** All results may automatically be reported to DOH if required by state.

Turnaround Time (TAT) Options - Please Check
 3 Hour 6 Hour 24 Hour 48 Hour 72 Hour 96 Hour 1 Week 2 Week

Microbiology Test Codes			
M001 Air-O-Cell	M174 MoldSnap	M012 Pseudomonas aeruginosa (PIA***)	M115 Sewage Screen - Water (PIA***)
M030 Micro 5	M032 Allergenco-D	M024 Pseudomonas aeruginosa (MFT*)	M116 Sewage Screen - Water (MPN**)
M041 Fungal Direct Examination		M015 Heterotrophic Plate Count	M117 Sewage Screen - Swab (PIA***)
M169 Pollen ID & Enumeration		M017 Total Coliform & E. coli (Colilert PIA***)	M013 Sewage Screen - Swab (MFT*)
M280 Dust Characterization Level-1		M018 Total Coliform & E. coli (MFT*)	M133 Methicillin-resistant Staph. aureus (MRSA)
M281 Dust Characterization Level-2		M114 Total Coliform & E. coli Enumeration (Colilert MPN**)	M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration
M005 Viable Fungi- Air Samples (Genus ID & Count)		M019 Fecal Coliform (MFT*)	M014 Endotoxin Analysis
M006 Viable Fungi- Air Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M020 Fecal Streptococcus (MFT*)	M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)
M007 Culturable fungi - Surface Samples (Genus ID & Count)		M029 Enterococci (MFT*)	Other See Analytical Price Guide
M008 Culturable fungi - Surface Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M129 Enterococci (Enterolert PIA***)	Legionella Analysis Please use EMSL Legionella COC
M009 Bacteria Culture Gram Stain & Count		M180 Real Time qPCR-ERMI 36 Panel	
M010 Bacteria Count & ID - 3 Most Prominent		M025 Sewage Screen -Water (MFT*)	
M011 Bacteria Count & ID - 5 Most Prominent			

*MFT= Membrane Filtration Technique
 **MPN= Most Probable Number
 ***PIA= Presence/Absence

Name of Sampler: Sheral Diaz **Signature of Sampler:** *[Signature]*

Sample #	Sample Location/Description	Sample Type	Potable/ NonPotable (Only for Waters)	Test Code	Volume/ Area	Date/Time Collected	Temperature (C) (Lab Use Only)
Example A1	Kitchen Sink/Tap	Water	<input checked="" type="checkbox"/> P <input type="checkbox"/> NP	M017	100 mL	9/1/13 4:00 PM	
S1	Cafeteria	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	25ml	12/04/20	
S2	HW between CR 11 and 12	"	<input type="checkbox"/> P <input type="checkbox"/> NP	"	"	"	
S3	Infront of CR 6A	"	<input type="checkbox"/> P <input type="checkbox"/> NP	"	"	"	
S4	HW between CR 05 and 06	"	<input type="checkbox"/> P <input type="checkbox"/> NP	"	"	"	
S5	Room Conference and planning	"	<input type="checkbox"/> P <input type="checkbox"/> NP	"	"	"	

Client Sample # (s): - **Total # of Samples:** 07 **Samples Received Chilled?** Yes / No (Lab Use Only)

Relinquished (Client): *[Signature]* **Date:** **Time:**
Received (Lab): *[Signature]* **Date:** **Time:**



Comments/Special Instructions:

RECEIVED
 EMSL ANALYTICAL, INC.
 BELTSVILLE, MD
 2020 DEC -4 PM 1:06



EMSL Analytical, Inc.

Sample Transfer Form

Receiving Lab:	EMSL- BELTSVILLE	Phone Number:	3019375700	
		Fax Number:	3019375701	
Relinquished to:	EMSL- PLYMOUTH MEETING	Phone Number:	8002203675	
		Fax Number:	8567860262	
Does new lab hold equivalent or additional accreditation? *			<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No	
EMSL Customer ID # (if known):	SALU50			
Client Name:	SALUT INC			
Client Project:	19-035/JOHN EDGAR HOWARD			
Tests to be Performed:	M001			
Date Received:	12/4/20			
Date Relinquished:	12/7/20			
Date Due:	3 DAYS - DUE 12/9 @ 1:06 PM			
Special Instructions: (e.g. Work Order # , required qualifications, project specific procedures/modifications)				
Relinquished by (Signature): 	Date: 12/7/20	Received by (Signature): 	Date: 12 8 20 1:40	
Relinquished by (Signature):	Date:	Received by (Signature):	Date:	
Customer Agreement - Please sign form and send to the receiving laboratory. By signing below, you agree to permit the above named receiving lab to transfer samples to a separate EMSL lab with equivalent qualifications* for analysis. The final report will be issued from the analyzing laboratory. Ensure any requirements are listed in special instructions.				
Name (please print):	Signature:	Agent of:	Date:	
<i>If this is a recurring project or sample type that may require samples to be relinquished on a regular basis, a Standing Agreement form must be completed.</i>				

* Receiving and analyzing labs shall be aware of required qualifications of project prior to transfer of samples.

Note: If customer has been notified and approved this transfer verbally or by e-mail, the receiving lab must sign for the customer above. EMSL employee filling out form on behalf of customer shall print name of person to whom they spoke, date agreement was received, and then sign under Signature.